

## Dr. Tessem Research

### Introduction

There are two major iterations of diabetes. Type 1 diabetes (T1D) is caused by autoimmune destruction of an individual's  $\beta$ -cells. An individual with T1D has severe insulin deficiency, and today is treated with multiple daily insulin injections. Type 2 diabetes (T2D) is associated with obesity and a sedentary lifestyle, and is characterized by insulin insensitivity in the muscle and adipose tissue. These patients are initially treated with insulin sensitizing compounds. While  $\beta$ -cell mass is originally maintained in T2D individuals, ultimately these patients lose  $\beta$ -cells and eventually also receive daily insulin injections. Therefore, T1D and T2D are ultimately characterized by decreased functional  $\beta$ -cell mass. Functional  $\beta$ -cell mass refers to the  $\beta$ -cell insulin secretion rate, the  $\beta$ -cell proliferation rate and the  $\beta$ -cell death rate. An increase in the  $\beta$ -cell secretion rate or proliferation, or a decrease in the cell death rate would all increase the functional  $\beta$ -cell mass. The ability to increase functional  $\beta$ -cell mass could be used as a tool to cure individuals with Type 1 and Type 2 diabetes.

The Tessem lab (Diabetes research group: [tessemlab.byu.edu](http://tessemlab.byu.edu)) is interested in delineating the molecular pathways that control functional  $\beta$ -cell mass with the ultimate application of increasing  $\beta$ -cells *ex vivo* for islet transplantation or *in situ* for enhancement of a patient's own residual  $\beta$ -cells. Our studies are focused on identifying the molecular accelerators and brakes that control  $\beta$ -cell replication, determining how these factors regulate functional  $\beta$ -cell mass, develop small molecule activators of the  $\beta$ -cell proliferation pathways and apply these findings to two state models of  $\beta$ -cell function such as obese vs. lean or aged vs. young models. Finally, our studies focus on discovering unique regulators of integrative metabolism.

Our experimental models use tissue culture  $\beta$ -cell lines, primary rodent islets and primary human islets. We use microarray expression analysis to determine genes involved in the particular pathways, followed by a variety of molecular and pharmacological techniques to manipulate pathways in order to understand how they effect functional  $\beta$ -cell mass. We measure changes in proliferation rate, glucose stimulated insulin secretion and protection against apoptosis as metrics of  $\beta$ -cell function.

### Major Emphasis: Type I; Minor Emphasis: Type II

#### 1) Current research efforts

- a) Map molecular pathways that control  $\beta$ -cell proliferation, insulin secretion and protection from cellular death.
- b) Determine genes upstream of Nkx6.1 critical for functional  $\beta$ -cell mass
- c) Verify genes downstream of Nkx6.1 that are necessary and sufficient for functional  $\beta$ -cell mass

- d) Using small molecule chemistry find agonist that manipulate key players in the functional  $\beta$ -cell mass pathway (i.e. Nr4a1, Nr4a3)
- e) Explore the idea of  $\beta$ -cell subtypes in terms of proliferation, insulin secretion and protection from cell death

2) **Anticipated Research Results**

- a) Rescue Type 1 and Type 2 diabetic patients through increased islet mass
  - i) Ex vivo expansion of  $\beta$ -cell mass for transplantation
  - ii) In situ expansion of endogenous  $\beta$ -cells
- b) Classify subgroups of  $\beta$ -cells
- c) Explore function of Nr4a family on mitochondrial function
- d) Characterize the genes essential for the Nkx6.1  $\beta$ -cell proliferation and function pathway
  - i) Nkx6.1—  $\beta$ -cell specific transcription factor
  - ii) NKX6.1 activates VGF that protects against apoptosis and increases glucose stimulated insulin secretion
  - iii) Nkx6.1 activates Nr4a1 and Nr4a3 which stimulate beta cell proliferation

\*\*See full profile and publications [here](#)